

2,2-Disubstituted Analogues of the Natural Hormone 1α ,25-Dihydroxyvitamin D₃: Chemistry and Biology

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Abstract—Six new 2,2-disubstituted analogues of the natural hormone calcitriol have been prepared. Chemical novelty includes (1) the first example of an inverse-electron-demand Diels—Alder cycloaddition using a pyrone diene and a difluorinated vinyl ether dienophile, leading to difluorinated analogues 7 and (2) a conceptually streamlined approach to dimethylated 19-nor analogues 8. Analogues 7a and 8a are similar to calcitriol in terms of in vitro antiproliferative activity, but they are different from calcitriol in terms of transcriptional activity: difluorinated analogue 7a is 2–3 times more active transcriptionally than calcitriol, whereas dimethylated analogue 8a is 7.5 times less active transcriptionally. Whereas the in vivo calcemic activity of difluorinated analogue 7a is similar to that of calcitriol, dimethylated analogue 8a is considerably less calcemic than calcitriol. Dimethylated analogue 8a strongly suppresses parathyroid hormone (PTH) secretion. © 2002 Elsevier Science Ltd. All rights reserved.

Introduction

The vitamin D family of steroid hormones is being studied by diverse scientists interested in issues ranging from fundamental organic and medicinal chemistry to molecular biology, endocrinology, and new drug development. Much of the motivation for this widespread interest and effort is the growing appreciation that the natural hormone 1α,25-dihydroxyvitamin D₃ (calcitriol, 1) has many different human biological functions ranging from regulating calcium and phosphorus homeostasis to governing cell growth and differentiation.² Medical use of the natural hormone calcitriol (1), however, is often risky due to the hypercalcemia typically induced by the supraphysiological levels of this hormone required to treat humans with diseases such as cancer, psoriasis, and autoimmune disorders.^{1,2} Small chemical changes at various positions in the structure of the natural hormone have produced thousands of synthetic analogues, some of which have desirably low calcemic activity and high antiproliferative and pro-

Structural changes specifically at the 2-position of the natural hormone in several cases produce analogues having desirable physiological properties. For example, the Chugai pharmaceutical company designed some 2-alkoxy and 2-alkyl analogues (e.g., **2** and **3**), among which ED-71 (**2**) is a leading drug candidate to treat women who have osteoporosis.⁴ Also, we^{5,6} and others^{7–9} designed a series of 2-alkyl analogues (**4–6**) and showed that some of these 2-monosubstituted analogues have strong affinity for the vitamin D receptor (VDR). Additionally, 2 β -fluoro-19-norcalcitriol¹⁰ and 2 β -fluorocalcitriol has been prepared, and 4,4-difluorocalcitriol has been prepared to probe A-ring conformation. No 2,2-disubstituted analogue of calcitriol (**1**), however, has been reported. Now, we describe a series of six 2,2-disubstituted analogues, prepared using

differentiation activities.³ Several of such designer analogues are now undergoing preclinical evaluation for chemotherapy of diverse human diseases, and some of these synthetic analogues are U.S. FDA-approved drugs (e.g. calcipotriol for treatment of psoriasis and $1\alpha,25$ -dihydroxy-19-nor-vitamin D_2 for treatment of secondary hyperparathyroidism).²

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some novel organic chemistry. Strong electronic influence by two powerfully electronegative fluorine atoms¹⁴ at position-2 was expected to enhance the hydrogenbonding ability of the 1- and 3-OH groups and to afford new analogues 7 with enhanced VDR affinity and lipophilicity. Considerable steric hindrance (but minimal electronic influence) by two methyl groups at position-2 was expected to diminish the reactivity of the 1- and 3-OH groups (e.g., retarding 3-epimerization and A-ring dehydration)¹⁵ in new 19-nor analogues 8 but probably also to interfere with VDR binding.

Chemistry

We have extensively developed Diels-Alder cycloaddition reactions of heteroaromatic 2-pyrones with electronically matched dienophiles to form synthetically versatile and stereochemically rich bicyclic lactone adducts. ¹⁶ We report here the first example of a successful high pressure 4+2-cycloaddition between commercial electron-poor 3-methoxycarbonyl-2-pyrone and a new geminally difluorinated electron-rich vinyl ether dienophile (Scheme 1); chromatographically pure racemic cycloadduct 9 was isolated in 73% yield, formed regiospecifically and stereospecifically with the desired 1,3-trans dioxygenation pattern necessary for ultimate transformation into calcitriol analogues 7. Nucleophilic opening of the lactone in 9 using lithium allyloxide¹⁷

proceeded smoothly to form polyfunctionalized cyclohexene 10 without intermolecular or intramolecular displacement of fluoride ion. Also, no loss of fluoride was encountered during the several subsequent steps in Scheme 1 using powerful nucleophiles (e.g., LiAlH₄, Ph₂PLi) or using high temperature for Claisen rearrangement of the sulfoxide-containing vinyl ether derived from allylic alcohol 12; conjugated dienyl ester 13 was formed after in situ sulfoxide pyrolytic elimination.¹⁸ Standard protocol¹⁹ allowed smooth conversion of dienyl ester 13 into racemic 2,2-difluorinated A-ring phosphine oxide 14. Coupling of phosphine oxide (\pm) -14 with enantiomerically pure C,D-ring ketone (+)-15, followed by desilylation, produced the desired 2,2difluorinated analogues (-)-7a and 7b in a 5:1 ratio. Distinction between these two diastereomers was based, as in previous related cases, on their characteristic ¹H NMR (Table 1).¹⁹ Based on chromatographic comparison and as expected based on the presence of two fluorine atoms, difluorinated analogue 7a is noticeably more lipophilic than calcitriol (1).

To simplify preparation of some 2,2-dimethyl analogues of calcitriol (1), we chose to design analogues lacking the 19-methylene group; DeLuca's pioneering work has shown that many 19-nor analogues of calcitriol (1) retain much of the hormone's desirable cell growth regulatory properties while having low calcemic activities.²⁰ Our streamlined synthetic approach, outlined in

$$F_{3}C \cap OH \xrightarrow{II. \ PMBCI, \ n \cdot Bu_{4} \text{NI } (cat) \ THF : DMPU} \\ \hline PBULI \ Et_{2}O:\text{hexanes} \\ \hline 32\% \\ \hline O \ CO_{2}Me \ F \ OPMB \ CH_{2}Cl_{2} + FO_{2}Me \ OPMB \$$

Scheme 1.

Table 1. ^{1}H NMR (δ) and optical rotation characteristics of new analogues

Analogue	C_{18} - CH_3	C ₆ -H	C ₇ -H	$[\alpha]_{\mathrm{D}}^{25}$
7a	0.55	6.45	6.02	
7b	0.54	6.47	6.00	
8a	0.54	6.29	5.84	+ 29
8b	0.55	6.30	5.83	+21
8c	0.54	6.31	5.85	+94
8d	0.55	6.33	5.86	+42

Scheme 2 starting with symmetrical 2,2-dimethyl-1,3cyclohexanedione, involves a series of intermediates grouped into two families: (1) a pair of diastereomeric trans-1,3-dihydroxycyclohexanes 16 and (2) the corresponding cis-1,3-dihydroxycyclohexane 17 having symmetrical substitution about a plane through C-2 and C-5 of the cyclohexane ring and, therefore, having also a simplified ¹H NMR spectrum especially in the δ 3.7 region (see Experimental). Noteworthy is the Michael addition of a phenylthioacetate enolate that occurs exclusively trans to the resident siloxy group leading to two 3-siloxy-5-phenylthioacetate cyclohexanone precursors to adducts 16 and 17. In these two cyclohexanone precursors to adducts 16 and 17, proton NMR shows clearly that the 3- and 5-substituents are *trans* to each other, with the 3-siloxy group axial and with the equatorial C-3 H appearing at δ 3.8 as a doublet of doublets with coupling constants of 2.2 and 4.0 Hz. Sulfide oxidation of thioether adducts 16 and 17 into the corresponding sulfoxides (as a mixture of diastereomers that were not separated) and then pyrolysis gave the pure α,β -unsaturated esters 18 and 19 in high yields. Standard transformations¹⁹ then produced the A-ring phosphine oxides **20** and **21**. Each of these racemic Aring units underwent separate Horner–Wadsworth–Emmons coupling with enantiomerically pure C,D-ring ketone (+)-**15** to form chromatographically separated 2,2-dimethyl-19-nor analogues **8a** and **8b** (in low yields) and also **8c** and **8d** (in good yields). Distinction between 1,3-*trans* diastereomers **8a** and **8b** and between 1,3-*cis* diastereomers **8c** and **8d** was based on optical rotations and ¹H NMR (Table 1)¹⁹ and also on analogy with analogues of calcitriol (**1**) having similar 1,3-diol stereochemical relationships.²¹

Biology

Motivation for interest in non-calcemic analogues of calcitriol (1) rests not only on their potential chemotherapeutic applications but also on using these analogues as sensitive probes of the fundamental molecular mechanisms underlying the varied biological effects that such analogues elicit. For example, we have discovered that there is an inverse relationship between some analogues' transcriptional potencies and their interaction specifically with AF-2 residues of VDR²² and that some A-ring analogues with low affinity for VDR modulate chondrocyte growth via membrane effects that are dependent on the stage of cell maturation.²³ New 2,2difluoro analogues 7 and 2,2-dimethyl analogues 8 now provide some surprising observations; understanding the molecular biology underlying these results may advance this field in significant ways.

The antiproliferative activity, determined in vitro using murine keratinocytes as previously described,²⁴ of only one of the four 2,2-dimethyl diastereomers (i.e., **8a** with

natural A-ring diol stereochemistry) is similar to that of calcitriol (1, Fig. 1). Likewise, 2,2-difluoro diastereomer 7a with natural A-ring diol stereochemistry has antiproliferative activity similar to that of calcitriol (1, Fig. 1). The VDR-mediated transcriptional potencies of 2,2disubstituted analogues 7a and 8a, determined in vitro using a previously described protocol in rat osteosarcoma ROS 17/2.8 cells,²⁵ are different from that of calcitriol (1). The ED₅₀ values for transcriptional activity are as follows: calcitriol (1), 0.4 nM; difluoro analogue 7a, 0.15 nM; dimethyl analogue 8a, 3.0 nM. Competition assays²⁵ using the recombinant human vitamin D receptor revealed the affinities of these two analogues [relative to 100% binding of calcitriol (1)] to be 9.6% for 7a and 1.3% for 8a. Thus, as rationally designed, the presence of two methyl groups at the 2-position in analogue 8a strongly diminishes (by a factor of 75) binding of this analogue to the human VDR.

Scheme 2.

Remarkably however, this 2,2-dimethyl analogue 8a is only 7.5-fold less active transcriptionally than calcitriol (1) in rat osteosarcoma cells. Likewise, although 2,2-difluoro analogue 7a binds to the human VDR only about 1/10 as well as the natural hormone 1, 7a is 2–3 times more transcriptionally active in the rat osteosarcoma cells than the natural hormone 1. Therefore, as in some related instances, ^{22,26} the levels of genomic activity produced by analogues 7a and 8a are not directly proportional to their affinities for the nuclear VDR.

2,2-Dimethyl analogue 8a with natural 1α ,3 β -diol stereochemistry (but not the diastereomeric 1α ,3 α -diol analogue 8c) is a strong inhibitor in vitro of parathyroid hormone (PTH) secretion (Fig. 2). Confluent cultures of bovine parathyroid cells were incubated for 72 h with various concentrations of dimethlylated analogues 8a, 8c or calcitriol (1). The steady state levels of PTH secretion were then determined during a 3-h incubation with fresh medium. As shown in Fig. 2, *trans* diol ana-

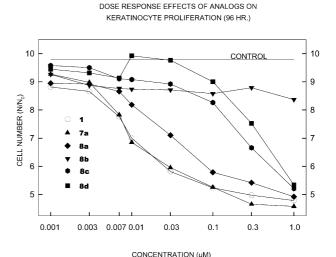


Figure 1.

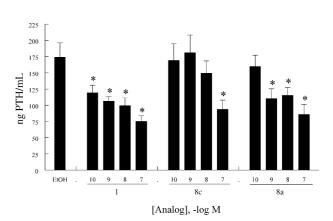


Figure 2. Suppression of PTH secretion. Confluent cultures of bovine parathyroid cells were treated for 72 h with the designated concentration of the vitamin D compounds. The cells were then incubated in fresh medium, and the amount of PTH secreted during a 3-h period was determined by RIA. The values are expressed as mean \pm SD (n=6). *p < 0.001.

logue 8a appeared to be about 10 times less active than calcitriol (1), whereas cis diol analogue 8c appeared to be several hundred times less active than calcitriol (1). Inhibition of PTH by calcitriol (1) is well established to involve repression of PTH gene transcription^{27,28} and vitamin D response elements (VDREs) have been identified in the promoters of the human,29 rat,30 and chick³¹ PTH genes. Further characterization of these negative VDREs and establishment of transcriptional assays have been hampered by the lack of a parathyroid cell line and the lack of activity of the PTH gene promoter in non-parathyroid cells. The suppression of PTH by vitamin D compounds is known to be VDR-dependent, but the other components of the negative regulatory complex have not been characterized. In fact, while VDR/RXR dimers appear to bind the rat and chick VDREs,^{30,31} the dimerization partner for VDR binding to the human PTH VDRE has not been identified.³² Thus, the molecular details of the mechanism for transcriptional repression by liganded VDR are not known. The VDR present in the parathyroid glands is functionally indistinguishable from that in other tissues. Studies examining structure–activity relationships for the suppression of PTH by vitamin D analogues have revealed a close correlation between the VDR affinity and suppressive activity. 33 The suppressive activity of PTH by trans diol analogue 8a correlates well with the transcriptional potency of this analogue in the rat osteosarcoma cells, further substantiating that this suppressive activity is mediated by the vitamin D receptor.

The in vivo calcemic activities of difluoro analogue 7a and dimethyl analogue 8a were determined using our standard protocol¹⁹ in which rats are treated daily with calcitriol (1) at $0.5 \, \mu g/kg$ body weight and with our new analogues at either $10.0 \, \mu g/kg$ body weight (Fig. 3) or

EFFECT OF VITAMIN D₃ ANALOGS ON CALCIUM LEVELS IN RAT URINE

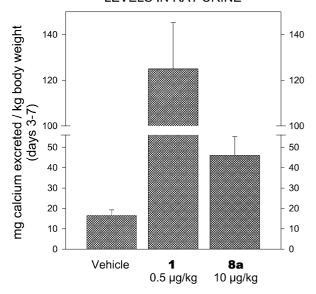


Figure 3. Effects of vitamin D_3 analogues on urinary calcium excretion in rats. Animals were treated with 0.5–10.0 microgram/kg body weight of test compound po for 7 consecutive days, and urinary excretion of calcium was measured during days 3–7. Values are mean + SE from three animals in each group.

1.0 μg/kg body weight (Fig. 4) daily for 1 week. At a 20-fold higher dose than calcitriol (1), dimethyl analogue **8a** caused only slight elevation of urinary calcium levels (Fig. 3) but no slowing of animal weight gain (data not shown). In sharp contrast, even at only a 2-fold higher dose than calcitriol (1), difluoro analogue **7a** elevated urinary calcium levels more than did calcitriol (1, Fig. 4). This strong calcemic activity of difluoro analogue **7a** was not expected based on its relatively low VDR binding affinity (9.6% that of the natural hormone 1) or based on the low calcemic effect of the closely related 2β-fluorocalcitriol. 12

In conclusion, rationally designed and newly synthesized 2,2-disubstituted chemical entities 7a and 8a illustrate how even very small structural changes in large molecules can produce powerful and different biological effects. Specifically, the desirable characteristics of high antiproliferative and PTH-suppression activities with low calcemic activity make new dimethylated analogue 8a suitable for further in vivo testing as a potential chemotherapeutic drug candidate. The new knowledge presented here about the relationships between chemical structure and biological activity may allow also other analogues of calcitriol (1) to be prepared for medicinal use and/or for use as sensitive probes of vitamin D molecular biology and mechanism of action. 34-36

Experimental¹⁹

p-Methoxybenzyl 2,2,2-trifluoroethyl ether. Trifluoroethanol (10 g, 0.10 mol) in THF (10 mL) was added dropwise via addition funnel to a stirred suspension of NaH (2.9 g, 0.12 mol) in THF (10 mL) at 0 °C over a period of 30 min. To the resulting mixture was added a

EFFECT OF VITAMIN D₃ ANALOGS ON CALCIUM LEVELS IN RAT URINE

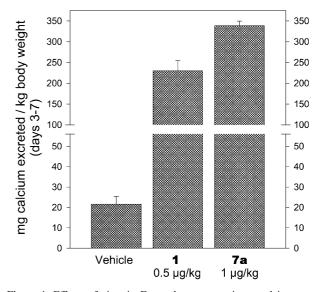


Figure 4. Effects of vitamin D_3 analogues on urinary calcium excretion in rats. Animals were treated with 0.5–10.0 microgram/kg body weight of test compound po for 7 consecutive days, and urinary excretion of calcium was measured during days 3–7. Values are mean + SE from three animals in each group.

solution of p-methoxybenzyl chloride (PMB-Cl, 19 g, 0.12 mol) in THF (10 mL) and N,N'-dimethylpropyleneurea (DMPU, 6 mL) via addition funnel over 1 h. After this addition, tetrabutylammonium iodide (n-Bu₄NI, 11 g, 0.03 mol) was added and the mixture warmed slowly to rt overnight. After acidic workup, the crude product was vacuum distilled to afford the desired ether as a pale orange liquid (25 g, quantitative): bp 100-101 °C/0.5 mm Hg; ¹H NMR (CDCl₃) δ 7.30–7.21 (m, 2H), 6.93-6.85 (m, 2H), 4.58 (s, 2H), 3.78 (s, 3H), 3.76 $(q, J=9.0 \text{ Hz}, 2H); ^{13}C \text{ NMR} (CDCl_3) \delta 159.7, 129.6,$ 128.5, 124.1 (q, J = 278.0 Hz), 113.9, 73.6, 66.6 (q, J = 33.7 Hz), 55.1; ¹⁹F NMR (CDCl₃, CFCl₃) $\delta - 74.2 \text{ to}$ -74.4 (m); IR (neat) 2937, 2837, 1613, 1515, 1280, 1250, 1164, 1117, 1034, 998, 965, 827, 665. Anal. Calcd for C₁₇H₁₆F₂O₆: C, 54.55; H, 5.04, found: C, 54.51; H, 5.24.

p-Methoxybenzyl 2,2-difluorovinyl ether. A solution of n-BuLi (26 mL, 0.26 mol, 10 M in hexanes) was added dropwise via addition funnel to a 1 L flask fitted with an argon inlet and outlet bubbler charged with a cold $(-78 \,{}^{\circ}\text{C})$, stirred solution of the aforementioned ether (23 g, 0.10 mol) in Et₂O (210 mL). The resulting solution was allowed to reach rt gradually overnight. Upon cooling the mixture to 0 °C, the reaction was quenched with dropwise addition of saturated aq NH₄Cl (50 mL). The reaction mixture was then extracted with Et₂O (3×50 mL) and the combined organics filtered through Celite, washed with H_2O (2×250 mL), dried (Na₂SO₄), filtered, and carefully concentrated to a brown oil. Vacuum distillation gave the desired 2,2-difluorovinyl ether as a pale yellow, somewhat unstable liquid (6.6 g, 32%) followed by a mixture of starting material and desired product (3.4 g, \sim 1:1). PMB-2,2-difluorovinyl ether: bp 98 °C/0.5 mm Hg; ¹H NMR (300 MHz, CDCl₃) δ 7.33–7.24 (m, 2H), 6.94–6.87 (m, 2H), 5.65 (dd, J = 22.0, 3.8 Hz, 1H), 4.67 (s, 2H), 3.82 (s, 3H). Due to its instability, this 2,2-difluorovinyl ether was used immediately.

4-Carbomethoxy-5-endo-[(p-methoxybenzyl)oxy]-6,6-difluoro-3-oxo-2-oxabicyclo-[2.2.2]oct-7-ene (\pm)-9. A fiveinch length of heat shrinkable Teflon tubing (Ace Glass #12685-40) was heat sealed at one end with a stainless steel plug. To the tube were added 3-carbomethoxy-2pyrone (1.0 g, 6.5 mmol), CH₂Cl₂ (10 mL), and PMB-2,2-difluorovinyl ether (6.5 g, 32 mmol). The tube was sealed at the other end with a stainless steel plug and placed under high pressure (10–12 kbar) for 5 days.³⁷ The tube was opened, the reaction mixture was transferred to a flask and concentrated in vacuo, and the residue purified by silica gel chromatography (5-40% EtOAc/hexanes). The fractions containing the desired syn,endo isomer (\pm) -9 were combined and concentrated to a faint yellow oil, which afforded white crystals on standing (1.7 g, 74%): mp 74–76°C; ¹H NMR (CDCl₃) δ 7.23–7.17 (m, 2H), 6.96–6.92 (m, 1H), 6.91–6.85 (m, 2H), 6.57–6.49 (m, 1H), 5.17–5.09 (m, 1H), 4.72 (d, J = 11.2 Hz, 1H), 4.49 (d, J = 11.2 Hz, 1H), 4.36 (d, J = 8.0 Hz, 1H), 3.80 (s, 3H), 3.79 (s, 3H); ¹³C NMR (CDCl₃) δ 165.5, 165.1, 159.7, 132.5, 130.3, 127.7, 126.6 (d, J = 5.3 Hz), 118.6 (t, J = 259.6 Hz), 113.7, 75.3 (t, J=8.7 Hz), 74.9 (t, J=8.4 Hz), 74.7, 74.2 (d, J=2.2

Hz), 61.0 (d, J=6.1 Hz), 55.2, 53.2; ¹⁹F NMR (CDCl₃, CFCl₃) δ-103.3 (ddd, J=244.9, 15.8, 5.1 Hz), -113.8 (d, J=244.9 Hz); IR (neat) 2954, 1780, 1751, 1613, 1515, 1340, 1295, 1250, 1171, 1131, 1103, 1049, 826. Anal. Calcd for C₁₇H₁₆F₂O₆: C, 57.63; H, 4.55, found: C, 57.64; H, 4.62.

Bis-silylated mixed malonate (\pm)-10. A solution of *n*-BuLi (1.6 M in hexanes, 0.900 mL, 1.44 mmol) was added dropwise to freshly distilled allyl alcohol (5 mL) at 0 °C. Bicyclic lactone (\pm)-9 (300 mg, 0.847 mmol) was then added in solution (5 mL CH₂Cl₂, 2.5 mL allyl alcohol) over 10 min. The reaction was stirred at 0 °C for 3 h and quenched with saturated aq NH₄Cl. The aqueous phase was extracted with EtOAc (3×25 mL), the combined organics dried over MgSO₄, and the solvents removed in vacuo to provide crude hydroxy mixed malonate as a pale yellow oil requiring no further purification for the following deprotection.

To the crude hydroxy mixed malonate was added CH_2Cl_2 , H_2O (4.25 and 0.25 mL, respectively), and DDQ (297 mg, 1.31 mmol). Additional DDQ (297 mg) was added after 6 h of stirring to complete the deprotection. After 12 h, the reaction was cautiously quenched with saturated aq NaHCO₃, decanted into H_2O (50 mL), extracted with CH_2Cl_2 (3×20 mL), dried over Na_2SO_4 , filtered through silica (1 inch plug), and concentrated to an oil requiring no further purification for the following bis-silylation.

The crude diol was dissolved in CH₂Cl₂ (8.0 mL), cooled to -78 °C, and treated with 2,6-lutidine (195 μ L, 1.68 mmol) and TBSOTf (339 µL, 1.48 mmol). After stirring for 3 h, additional 2,6-lutidine and TBSOTf were added (same amounts as above). The reaction mixture was allowed to warm to 0 °C, at which time additional 2,6-lutidine and TBSOTf were added (same amounts as above) and the solution stirred at 0°C overnight. Additional 2,6-lutidine and TBSOTf were added ($3\times$ s, same amounts as above) to complete the reaction. After normal aqueous workup, the crude oil was passed though flash silica gel (5% EtOAc/hexanes) to afford (\pm)-10 as a pale yellow oil [323 mg, 92% from (\pm) -9]: ¹H NMR (CDCl₃) δ 6.02 (dq, J=10.4, 1.6 Hz, 1H), 5.85 (ddt, J = 17.2, 10.4, 5.7 Hz, 1H), 5.73 (ddd, J = 10.4, 6.4, 2.4 Hz, 1H), 5.25 (dddd, J = 10.4, 6.4, 1.6, 1.2 Hz, 2H), 5.05 (d, J = 11.6 Hz, 1H), 4.61 (apparent dddt, J = 29.5, 13.1, 5.7, 1.2 Hz, 2H), 4.63–4.54 (m, 2H), 3.74 (s, 3H), 0.90 (s, 9H), 0.84 (s, 9H), 0.15 (d, J=2.8Hz, 3H), 0.10 (s, 6H), 0.09 (s, 3H); $^{13}\text{C NMR (CDCl}_3)~\delta$ 167.2, 166.3, 131.2, 129.4 (d, J = 7.6 Hz), 123.6, 119.3 (t, J = 249.4 Hz), 118.6, 71.8 (dd, J = 24.7, 6.0 Hz), 66.4 (dd, J = 19.8, 5.4 Hz), 63.0 (d, J = 5.4 Hz), 53.1, 25.6, 18.2, 18.0, -4.5 (d, J=6.0 Hz), -4.9 (d, J=5.4Hz), -5.3; ¹⁹F NMR (CDCl₃, CFCl₃) δ -115.7 (d, J = 246.7 Hz), -124.9 (dd, J = 246.7, 15.2 Hz); IR (neat) 2954, 2931, 2858, 1749, 1255, 1221, 1200, 1180, 1135, 1111, 1040, 870, 838, 780; HRMS: calcd for $C_{24}H_{48}F_2O_6Si_2 + Na$ 453.2386, found 453.2386.

Methyl ester (\pm)-11. A mixture of (\pm)-10 (266 mg, 0.511 mmol), formic acid (24 μ L, 0.638 mmol), Et₃N (93

μL, 0.664 mmol), Ph₃P (27 mg, 0.102 mmol), and palladium (II) acetate (12 mg, 0.051 mmol) in 1,4-dioxane (1.6 mL) was heated to 100 °C in a sealed tube for 16 h. Upon cooling, the tube was carefully purged under argon and dilute HCl added (1 mL, 1 M ag). The contents of the tube were transferred to a separatory funnel and partitioned between H2O and CH2Cl2, extracted (3×10 mL CH₂Cl₂), dried (MgSO₄), filtered, concentrated, and passed through flash silica gel (5% EtOAc/hexanes) to give (\pm) -11 (210 mg, 94%) as a clear oil which solidified on standing: mp 42-43 °C; ¹H NMR (CDCl₃) δ 6.91 (dd, J = 4.8, 2.8 Hz, 1H), 4.69 (t, J = 6.4 Hz, 1H), 4.40–4.24 (m, 1H), 3.75 (s, 3H), 2.75– 2.60 (m, 1H), 2.45 (ddd, J = 19.4, 9.0, 2.8 Hz, 1H), 0.91(s, 9H), 0.85 (s, 9H), 0.17 (d, J=1.6 Hz, 3H), 0.14 (s, 9H)3H), 0.09 (s, 3H), 0.08 (s, 3H); 13 C NMR (CDCl₃) δ 165.6, 139.6, 130.5 (d, J=6.1 Hz), 119.3 (t, J=249.4Hz), 67.8 (dd, J = 87.9, 20.2 Hz), 64.8 (t, J = 20.2 Hz), 51.8, 34.0, 25.7, 18.14, 18.11, -4.9, -5.0, -5.1; ¹⁹F NMR (CDCl₃, CFCl₃) δ -121.6 (d, J=241.0 Hz),-130.4 (ddd, J = 241.0, 25.1, 6.8 Hz); IR (neat) 2954, 2930, 2858, 1725, 1258, 1177, 1123, 1082, 900, 838, 781; HRMS: calcd for $C_{20}H_{38}F_2O_4Si_2 + Na$ 459.2174, found 459.2177.

Allylic alcohol (\pm)-12. To a solution of (\pm)-11 (200 mg, 0.458 mmol) in CH₂Cl₂ (5 mL) at -78 °C was slowly added diisobutylaluminum hydride (DIBAL-H, 1.01 mL, 1.01 mmol, 1.0 M in PhCH₃). This mixture was allowed to warm to rt and stirred (1 h) until the reaction was complete by TLC analysis (20% EtOAc/hexanes). The reaction was quenched with aqueous sodium potassium tartrate (1 mL, 2 N), the mixture was extracted with CH_2Cl_2 (3×6 mL), the combined organic layers were washed with H₂O (4 mL), dried (Na₂SO₄), concentrated under reduced pressure, and passed through flash silica gel (5–10% EtOAc/hexanes) to afford (\pm)-12 (158 mg, 84%) as a white solid: mp 42–44 °C; ¹H NMR (CDCl₃) δ 5.67 (m, 1H), 4.47 (t, J=9.2 Hz, 1H), 4.24– 4.06 (m, 1H), 4.11 (s, 2H), 2.58–2.40 (m, 1H), 2.34–2.17 (m, 1H), 1.63 (br s, 1H), 0.91 (s, 9H), 0.89 (s, 9H), 0.15 (s, 6H), 0.10 (s, 3H), 0.08 (s, 3H); 13 C NMR (CDCl₃) δ 136.5 (m), 123.2, 120.1 (t, J = 247.5 Hz), 68.7 (t, J = 25.8Hz), 67.0 (t, J = 24.0 Hz), 63.8, 33.0 (m), 25.8, 25.6, 18.2, 18.1,-4.7, -4.9, -5.1; ¹⁹F NMR (CDCl₃, CFCl₃) δ -122.5 to-124.5 (m); IR (neat) 3601-3094 (br), 2954, 2930, 2859, 1472, 1463, 1256, 1186, 1126, 1102, 1080, 896, 878, 862, 837, 778. Anal. Calcd for C₂₀H₃₈F₂O₄Si₂: C, 55.84; H, 9.37, found: C, 56.10; H, 9.49.

Z-dienoate ester (±)-13. A 25-mL hydrolysis tube containing a solution of (±)-**12** (158 mg, 0.387 mmol), 1-phenylsulfinyl-2,2,2-triethoxyethane³⁸ (289 mg, 1.01 mmol) and 2,4,6-trimethylbenzoic acid (TMBA, 6.8 mg, 0.041 mmol) in CH₂Cl₂ (2 mL) was heated to 115 °C for 16 h. After cooling to rt, the solvent was removed in vacuo and the resulting light yellow oil was purified by flash silica gel chromatography (2–5% EtOAc/hexanes) to afford a mixture of Z- and E-dienoate esters (64 mg, \sim 2.2:1.0) and pure Z-dienoate ester (±)-**13** (104 mg). Separation of the remaining Z/E mixture using preparative plate chromatography (1000 μm, 5% EtOAc/hexanes) afforded pure E-dienoate ester (14 mg, 8%)

and pure (\pm)-13 (42 mg, 146 mg total, 79%) both as clear oils. (\pm)-13: ¹H NMR (CDCl₃) δ 5.72 (br s, 1H), 5.42 (t, J=1.6 Hz, 1H), 5.26 (br s, 1H), 4.59–4.47 (m, 1H), 4.21–4.04 (m, 1H), 4.13 (q, J=7.0 Hz, 2H), 2.53 (dt, J=13.6, 1.4 Hz, 1H), 2.33 (ddd, J=13.6, 4.8, 1.4 Hz, 1H), 1.23 (t, J=7.0 Hz, 3H), 0.92 (s, 9H), 0.88 (s, 9H), 0.11 (d, J=12.4 Hz, 6H), 0.10 (d, J=8.8 Hz, 6H); ¹³C NMR (CDCl₃) δ 165.4, 149.0, 141.1, 119.7, 119.6 (t, J=250.2 Hz), 115.9, 72.6 (t, J=23.5 Hz), 69.4 (t, J=25.8 Hz), 60.0, 41.4, 25.6, 25.5, 18.3, 18.0, 14.1, -5.0, -5.1, -5.3; ¹⁹F NMR (CDCl₃, CFCl₃) δ -120.7 (d, J=242.3 Hz), -123.7 (d, J=242.3 Hz); IR (neat) 2956, 2931, 2896, 2858, 1730, 1646, 1473, 1366, 1255, 1222, 1184, 1161, 1122, 1098, 1037, 1006, 940, 893, 838, 800; HRMS: calcd for C₂₃H₄₂F₂O₄Si₂ 476.2590, found 476.2587.

A-ring phosphine oxide (\pm)-14. To a solution of (\pm)-13 (104 mg, 0.218 mmol) in PhCH₃/CH₂Cl₂ (3.0 mL, 2:1) at $-78\,^{\circ}$ C was slowly added a solution of DIBAL-H (480 μ L, 1.0 M in PhCH₃, 0.480 mmol) via syringe. The reaction was maintained at $-78\,^{\circ}$ C (1 h), then slowly warmed to $-50\,^{\circ}$ C at which time the reaction was complete by TLC analysis (25% EtOAc/hexanes). The reaction was quenched with aqueous sodium potassium tartrate (1 mL, 2 N), HCl (1 mL, 1 M aqueous) and H₂O (1 mL), the organic layer was separated and the aq layer extracted with CH₂Cl₂ (3×6 mL), the combined organics dried (Na₂SO₄) and concentrated to give the desired allylic alcohol (103 mg) as a colorless oil which was pure enough to be carried forward directly.

To a cold $(-78 \,^{\circ}\text{C})$ solution of the crude allylic alcohol in THF (2.5 mL) was added a solution of n-BuLi (178 μL, 0.283 mmol, 1.53 M in hexanes) dropwise via syringe. The resulting solution was warmed (0 °C) for 30 min, cooled $(-20\,^{\circ}\text{C})$, and treated with a precooled solution of TsCl (62 mg, 0.327 mmol) in THF (0.5 mL). After stirring for 30 min at 0 °C, the reaction mixture was cooled (-78 °C) and treated with enough of a precooled solution of lithium diphenylphosphide [generated from the addition of n-BuLi (623 µL, 0.990 mmol, 1.53 M in hexanes) to a cold $(-78 \,^{\circ}\text{C})$ solution of Ph₂PH (172 μL, 0.990 mmol) in THF (2.6 mL)] until an orange color persisted for 5 min (\sim 2.1 mL). The reaction mixture was quenched with H₂O (three drops), warmed to rt, the solvent removed in vacuo, the residual oil dissolved in CH₂Cl₂ (3.5 mL), treated with hydrogen peroxide (2 mL, 5% aqueous H₂O₂), and stirred vigorously for 1 h. After general aqueous workup, the crude reaction mixture was passed through flash silica (20-50% EtOAc/hexanes) to afford 62 mg [46% from (\pm) -13] of (\pm)-14 as a clear oil: ¹H NMR (CDCl₃) δ 7.75–7.65 (m, 4H), 7.56–7.41 (m, 6H), 5.45 (apparent dd, J=14.6, 7.0 Hz, 1H), 5.36 (t, J = 1.6 Hz, 1H), 4.96 (br s, 1H), 4.42– 4.30 (m, 1H), 4.10–3.99 (m, 1H), 3.35 (dt, J=14.2, 8.4 Hz, 1H), 3.17 (dt, J = 15.6, 6.9 Hz, 1H), 2.45 (d, J = 14Hz, 1H), 2.31–2.21 (m, 1H), 0.91 (s, 9H), 0.81 (s, 9H), $0.10 \text{ (s, 3H)}, 0.07 \text{ (s, 3H)}, 0.05 \text{ (s, 3H)}, 0.02 \text{ (s, 3H)}; ^{13}\text{C}$ NMR (CDCl₃) δ 141.8 (d, J = 2.2 Hz), 137.8 (d, J = 11.4Hz), 133.1 (d, J = 33.4 Hz), 132.1 (d, J = 33.4 Hz), 131.9 (t, J=2.2 Hz), 130.9 (dd, J=9.1, 3.8 Hz), 128.6 (dd, J=11.4, 3.1 Hz), 120.0 (t, J=245.2 Hz), 117.5 (d, J=7.6 Hz), 114.9, 72.7 (t, J=22.3 Hz), 69.5 (t, J=26.5 Hz), 40.9, 31.5 (d, J=70.6 Hz), 25.62, 25.57, 18.3, 18.0, -5.0, -5.1; ¹⁹F NMR (CDCl₃, CFCl₃) δ -118.0 to -125.0 (m); IR (neat) 2952, 2929, 2856, 1472, 1438, 1255, 1166, 1120, 1093, 939, 892, 837, 780, 695, 552, 509; HRMS: calcd for $C_{33}H_{49}F_2O_3PSi_2$ 618.2926, found 619.3007.

General HWE Coupling: 2,2-difluorocalcitriol analogues (-)-7a and 7b. Prior to reaction, phosphine oxide and C,D-ring ketone (+)-15³⁹ azeotropically dried with benzene and left under vacuum for 48 h. To a solution of (\pm) -14 (63 mg, 0.102 mmol) in THF (1.1 mL) at -78 °C was added a solution of n-BuLi (64 μL, 0.101 mmol, 1.59 M in hexanes). After stirring in the dark for 1 h, a precooled (-78 °C) solution of (+)-15 (32 mg, 0.08 mmol) in THF (0.3 mL) was slowly cannulated into the flask containing the redorange ylide. The reaction mixture was maintained at -78 °C (3 h) then warmed to -40 °C (2 h). During this time the red-orange color faded to light vellow. The reaction was quenched with H₂O (3 mL), extracted with Et₂O (3×5 mL), dried (MgSO₄), filtered, and concentrated to give a crude product which was purified by flash silica gel chromatography (1% EtOAc/hexanes) to give a mixture of silyl protected analogues (23 mg, 57%), followed by recovered (+)-15 (elution with 20% EtOAc/hexanes, 7 mg, 37%) and (\pm) -14 (elution with 50% EtOAc/hexanes, 27 mg, 43%).

Half of the mixture of silyl-protected analogues was dissolved in THF (1.5 mL), treated with tetrabutylammonium fluoride (TBAF, 68 µL, 0.068 mmol, 1 M in THF), allowed to stir at rt overnight, and concentrated to an oil. Purification by flash silica gel chromatography (40% EtOAc/hexanes) gave 4.2 mg (58%) of a mixture of (-)-7a and 7b. The other half of this mixture was dissolved in EtOH (3.0 mL), cooled to 0°C, and treated with a solution of hydrogen fluoride (100 µL, 49% ag HF), warmed to room temperature over 2 h, and treated with additional HF solution (3×100 μL) to complete global deprotection. Upon completion by TLC analysis (50% EtOAc/hexanes), the reaction mixture was cautiously quenched with saturated NaHCO₃ solution and subjected to a general extraction. Purification by flash silica gel chromatography (40% EtOAc/hexanes) afforded 3.4 mg (47%) of the diastereomeric mixture. The combined mixture of diastereomers was subjected to HPLC separation (CHIRALCEL® OJ semipreparative column, 15% i-PrOH/hexanes, 3 mL/min) to afford only pure diastereomer 7a. (-)-7a: (3 mg, 12%, $1\alpha,3\beta$, R_f 37.2 min); [α]-12.6 (c 1.0, CHCl₃); ¹H NMR (CDCl₃) δ 6.45 (d, J=11.2 Hz, 1H), 6.02 (d, J=11.2 Hz, 1H), 5.62-5.55(m, 1H), 5.26–5.20 (m, 1H), 4.60–4.46 (m, 1H), 4.28– 4.15 (m, 1H), 2.81 (dd, J = 14.0, 4.8 Hz, 1H), 2.68 (d, J = 14.0 Hz, 1H), 2.45 (ddd, J = 14.0, 4.8, 3.8 Hz, 1H), 1.22 (s, 6H), 0.94 (d, J = 6.4 Hz, 3H), 0.55 (s, 3H); ¹³C NMR (CDCl₃) δ 144.8, 141.4, 129.0, 126.9, 118.1 (t, J = 247.0 Hz), 116.7, 115.7, 71.9 (t, J = 22.0 Hz), 71.1, 69.4–68.5 (m), 56.5, 56.4, 46.0, 44.4, 40.4, 39.3, 36.3, 36.1, 29.4, 29.2, 27.6, 23.7, 22.2, 20.8, 18.8, 12.0; ¹⁹F NMR (CDCl₃, CFCl₃) δ -121.6 to -126.0 (m); IR

(neat) 3636–3072 (br), 2937, 2869, 1377, 1215, 1156, 1077, 757; HRMS: calcd for $C_{27}H_{42}F_2O_{3+}Na$ 475.3000, found 475.3002.

5-(t-Butyldimethylsilyl)oxy-6,6-dimethyl-2-cyclohexenone. 3-Hydroxy-2,2-dimethylcyclohexanone (2.39 g, 16.8) mmol), prepared according to literature procedures, 40 was dissolved in CH₂Cl₂ (50 mL), cooled to -78 °C, and treated with 2,6-lutidine (2.92 mL, 25.2 mmol) and TBSOTf (3.84 mL, 16.8 mmol). The solution was stirred for 30 min at-78 °C, quenched with H₂O (30 mL), and extracted with CH₂Cl₂. The combined organics were dried over MgSO₄ and concentrated in vacuo. Silica gel chromatography (3% EtOAc/hexanes) provided 3-(tbutyldimethylsilyl)oxy-2,2-dimethylcyclohexanone as a colorless oil: ¹H NMR (CDCl₃) δ 3.64 (dd, J=7.2, 2.8 Hz, 1H), 2.34 (apparent t, J = 6.8 Hz, 2H), 2.04–1.85 (m, 2H), 1.77–1.66 (m, 1H), 1.65–1.53 (m, 1H) 1.08 (s, 3H), 1.04 (s, 3H), 0.84 (s, 9H), 0.01 (s, 3H), 0.00 (s, 3H); ¹³C NMR (CDCl₃) δ 214.6, 78.2, 51.4, 37.1, 29.2, 25.6, 23.1, 20.4. 17.9. -4.4. -5.2: IR (CHCl₃) 2954. 2858. 1713. 1472, 1255, 1120, 1082, 1002, 869, 835, 775; HRMS (EI) m/z (M⁺) calcd 256.1859 for C₁₄H₂₈O₂Si, found 256.1856.

3-(t-Butyldimethylsilyl)oxy-2,2-dimethylcyclohexanone (1.53 g, 5.96 mmol) in THF (15 mL) was cooled to -78 °C and then cannulated into a freshly prepared solution of LDA (6.55 mL, 1 M in THF/hexanes). The reaction mixture was stirred for 15 min and then cooled in liquid nitrogen until the THF just begins to form a slurry. A solution of PhSeCl (3.30 g, 17.2 mmol) in CH₂Cl₂ (15 mL) was added all at once. The reaction temperature increased to -55 °C upon addition. Dilute HCl (6.5 mL, 1 N aq) was added and the reaction mixture was warmed to rt, dried (MgSO₄), and the organic solvents removed in vacuo. The residue was taken up in CH₂Cl₂ (40 mL), the mixture cooled to 0 °C, and H₂O₂ (3 mL, 29% aqueous) was added over 1 h. The reaction mixture was dried over MgSO₄, the organic solvents removed in vacuo, and the residue purified by silica gel chromatography (3% EtOAc/hexanes) to provide the desired 5-(t-butyldimethylsilyl)oxy-6,6-dimethyl-2-cyclohexenone (757 mg, 50%) as a colorless oil: ¹H NMR (CDCl₃) δ 6.73 (ddd, J = 10.0, 5.2, 3.2 Hz, 1H), 5.96 (ddd, J=10.0, 2.2, 1.6 Hz, 1H), 3.83 (dd, J=7.6, 4.8 Hz,1H), 2.53 (ddt, J = 18.8, 4.8, 1.6 Hz, 1H), 2.39 (dddd, J = 18.8, 7.6, 3.0, 2.6 Hz, 1H, 1.12 (s, 3H), 1.05 (s, 3H),0.89 (s, 9H), 0.07 (s, 3H), 0.05 (s, 3H); ¹³C NMR $(CDCl_3)$ δ 204.1, 144.5, 128.4, 74.4, 48.6, 32.8, 25.7, 21.3, 18.2, 17.9, -4.3, -5.0; IR (CDCl₃) 2956, 2931, 2857, 1681, 1253, 1099, 876, 837, 776; HRMS (EI) m/z (M^+) calcd 254.1702 for $C_{14}H_{26}O_2Si$, found 254.1698.

1,3-trans Bis protected diols (\pm)-16. To ethyl phenylthioacetate⁴¹ (3.23 g, 16.5 mmol) in THF (30 mL) at $-78\,^{\circ}$ C was added a freshly prepared solution of LDA (15.0 mL, 1 M in THF/hexanes) and the reaction mixture was stirred for 30 min. A solution of 5-(*t*-butyl-dimethylsilyl)oxy-6,6-dimethyl-2-cyclohexenone (952 mg, 3.74 mmol) in THF (30 mL) was added, the reaction was warmed to $-30\,^{\circ}$ C, stirred for 12 h, quenched with H₂O (30 mL), and extracted with CH₂Cl₂. The combined organics were washed once with brine and dried

over MgSO₄, then concentrated in vacuo. Silica gel chromatography of the residue (3% EtOAc/hexanes) provided two diastereomeric thioethers (609 mg, 36%; 504 mg, 30%). Less polar thioether: ¹H NMR (CDCl₃) δ 7.48–7.24 (m, 5H), 4.22–4.05 (m, 2H), 3.82 (dd, J=4.2, 2.2 Hz, 1H), 3.57 (d, J=7.6 Hz, 1H), 2.79 (ddddd, J=12.4, 12.4, 7.6, 4.8, 3.8 Hz, 1H), 2.70 (ddd,J = 14.6, 4.8, 2.0 Hz, 1H), 2.44 (dd, J = 14.6, 12.4 Hz, 1H), 2.09 (ddd, J = 13.6, 12.4, 2.2 Hz, 1H), 1.83 (dddd, J = 13.6, 4.2, 3.8, 2.0 Hz, 1H), 1.20 (t, J = 7.2 Hz, 3H), 1.13 (s, 3H), 1.07 (s, 3H), 0.83 (s, 9H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C NMR (CDCl₃) δ 213.0, 171.1, 134.1, 132.2, 129.1, 127.8, 77.7, 61.4, 57.4, 50.1, 41.0, 34.8, 31.8, 25.8, 24.3, 21.3, 18.0, 14.0, -4.5, -5.1; IR (CHCl₃) 2956, 2930, 2857, 1732, 1714, 1472, 1258, 1151, 1063, 1025, 832, 776; HRMS (EI) m/z (M+H+) calcd 451.2338 for C₂₄H₃₈O₄SSi, found 451.2343. More polar thioether: ¹H NMR (CDCl₃) δ 7.47–7.23 (m, 5H), 4.19–4.06 (m, 2H), 3.84 (dd, J = 3.8, 2.2 Hz, 1H), 3.62 (d, J = 6.8 Hz, 1H), 2.83 (ddddd, J = 12.4, 12.0, 6.8, 4.8, 4.0 Hz, 1H), 2.68 (dd, J = 14.4, 12.4 Hz, 1H), 2.37 (ddd, J = 14.4, 4.8, 2.0 Hz,1H), 2.12 (dddd, J = 14.0, 4.0, 3.8, 2.0 Hz, 1H), 2.01 (ddd, J = 14.0, 12.0, 2.2 Hz, 1H), 1.19 (t, J = 7.2 Hz, 3H), 1.14 (s, 3H), 1.07 (s, 3H), 0.85 (s, 9H), 0.06 (s, 3H), 0.04 (s, 3H); ¹³C NMR (CDCl₃) δ 212.9, 171.0, 133.6, 132.6, 129.0, 127.9, 77.7, 61.3, 57.0, 50.2, 40.9, 34.7, 32.4, 25.7, 24.3, 21.3, 18.0, 14.1, -4.5, -5.1; IR (CHCl₃) 2956, 2930, 2857, 1732, 1714, 1472, 1257, 1154, 1102, 1063, 1027, 833, 776; HRMS (EI) m/z (M+H+) calcd 451.2338 for C₂₄H₃₈O₄SSi, found 451.2338.

To a solution of the less polar thioether (585 mg, 1.30 mmol) in MeOH (40 mL) was added NaBH₄ (5 mg, 1.5H⁻ equiv) in portions until starting material was consumed. The reaction was quenched with H₂O and extracted with EtOAc. The combined organics were washed with brine and dried over MgSO₄. After removal of the organic solvents in vacuo, silica gel chromatography provided two diastereomeric alcohols (162 mg, 27%; 362 mg, 62%) as colorless oils. Less polar 1,3-cis alcohol: ¹H NMR (CDCl₃) δ 7.47–7.22 (m, 5H), 4.19-4.02 (m, 2H), 3.95 (d, J=10 Hz, 1H), 3.70-3.66 (m, 1H), 3.56 (d, J=8.0 Hz, 1H), 3.55–3.50 (m, 1H), 2.66-2.54 (m, 1H), 2.26-2.18 (m, 1H), 1.86-1.58 (m, 3H), 1.18 (t, J = 7.2 Hz, 3H), 1.14 (s, 3H), 0.89 (s, 9H), 0.88 (s, 3H), 0.08 (s, 3H), 0.07 (s, 3H); ¹³C NMR (CDCl₃) δ 171.5, 134.0, 132.5, 128.9, 127.6, 77.5, 75.7, 61.0, 57.2, 37.6, 33.4, 32.4, 28.1, 25.8, 24.7, 24.4, 17.8, 14.1, -4.7, -5.2; IR (CHCl₃) 3511 (br), 2954, 2930, 2858, 1732, 1472, 1258, 1151, 1094, 1052, 838, 778; HRMS (EI) m/z (M+H+) calcd 453.2495 for C₂₄H₄₀O₄SSi, found 453.2494. More polar 1,3-trans alcohol: ¹H NMR (CDCl₃) δ 7.46–7.20 (m, 5H), 4.19– 4.00 (m, 2H), 3.80 (dd, J = 11.4, 4.6 Hz, 1H), 3.59 - 3.55(m, 1H), 3.52 (d, J = 8.8 Hz, 1H), 2.47–2.35 (m, 1H), 2.26-2.18 (m, 1H), 1.68-1.50 (m, 3H), 1.17 (t, J=7.2Hz, 3H), 1.03 (s, 3H), 0.88 (s, 9H), 0.82 (s, 3H), 0.03 (s, 3H), 0.01 (s, 3H); ¹³C NMR (CDCl₃) δ 171.5, 134.1, 132.3, 128.9, 127.5, 76.6, 72.5, 61.0, 57.4, 40.0, 34.4, 33.0, 32.9, 25.8, 24.4, 18.1, 18.0, 14.0, -4.5, -5.1; IR (CHCl₃) 3426 (br), 2954, 2930, 2857, 1732, 1472, 1256, 1150, 1070, 833, 775; HRMS (EI) m/z (M+H⁺) calcd 453.2495 for C₂₄H₄₀O₄SSi, found 453.2503.

Likewise, the more polar thioether (504 mg, 1.12 mmol), MeOH (30 mL), and NaBH₄ (13 mg, 1.5H⁻ equiv) provided two diastereomeric alcohols (257 mg, 51%; 162 mg, 31%) as colorless oils. Less polar, 1,3-cis alcohol: ¹H NMR (CDCl₃) δ 7.46–7.22 (m, 5H), 4.19–4.05 (m, 2H), 3.72-3.68 (m, 1H), 3.61 (d, J=6.8 Hz, 1H), 3.53-3.49 (m, 1H), 2.69-2.57 (m, 1H), 2.10-2.02 (m, 1H), 1.91–1.79 (m, 2H), 1.75–1.66 (m, 1H), 1.26 (t, J=7.2Hz, 3H), 1.14 (s, 3H), 0.91 (s, 9H), 0.90 (s, 3H), 0.10 (s, 3H), 0.09 (s, 3H); ¹³C NMR (CDCl₃) δ 171.5, 134.7, 131.9, 128.9, 127.4, 77.5, 75.7, 61.1, 57.9, 37.5, 33.5, 32.2, 28.3, 25.7, 24.6, 24.3, 17.8, 14.0, -4.7, -5.3; IR (CHCl₃) 3512, 2954, 2930, 2858, 1733, 1472, 1257, 1150, 1095, 1052, 837, 777; HRMS (EI) m/z (M+H⁺) calcd 453.2495 for C₂₄H₄₀O₄SSi, found 453.2489. More polar 1,3-trans alcohol: ¹H NMR (CDCl₃) δ 7.46–7.21 (m, 5H), 4.17-4.04 (m, 2H), 3.79 (dd, J=11.6, 4.8 Hz, 1H), 3.62-3.58 (m, 1H), 3.55 (d, J=8.0 Hz, 1H), 2.49-2.37(m, 1H), 1.98–1.91 (m, 1H), 1.83–1.76 (m, 1H), 1.58– 1.38 (m, 3H), 1.17 (t, J = 7.2 Hz, 3H), 1.04 (s, 3H), 0.89 (s, 9H), 0.83 (s, 3H), 0.06 (s, 3H), 0.04 (s, 3H); ¹³C NMR (CDCl₃) δ 171.7, 134.6, 132.0, 128.9, 127.4, 76.6, 72.5, 61.1, 57.7, 40.0, 34.4, 33.3, 32.6, 25.9, 24.4, 18.1, 18.0, 14.0, -4.4, -5.1; IR (CHCl₃) 3428 (br), 2954, 2929, 2857, 1733, 1472, 1255, 1153, 1071, 980, 832, 775; HRMS (EI) m/z (M+H⁺) calcd 453.2495 for C₂₄H₄₀O₄SSi, found 453.2585.

Each of the 1,3-trans alcohols from above was protected as follows: to a solution of alcohol (1 equiv) in CH₂Cl₂ at -30°C were added 2,6-lutidine (1.25 equiv) and TBSOTf (1.1 equiv). The reaction mixture was stirred for 30 min, quenched with H₂O, and extracted with CH₂Cl₂. The combined organics were dried over MgSO₄, the solvents removed in vacuo, and the residue purified by column chromatography to provide two 1,3trans bis silyl ethers (\pm) -16 both as a colorless oils. First 1,3-trans bis silyl ether (197 mg, 347 μmol, 97%); ¹H NMR (CDCl₃) δ 7.47–7.20 (m, 5H), 4.20–4.10 (m, 1H), 4.09-3.99 (m, 1H), 3.75 (dd, J=7.2, 4.4 Hz, 1H), 3.56-3.53 (m, 1H), 3.48 (d, J = 8.0 Hz, 1H), 2.42–2.29 (m, 1H), 2.11-2.02 (m, 1H), 1.66-1.56 (m, 1H), 1.52-1.43 (m, 1H), 1.29-1.18 (m, 1H), 1.18 (t, J=7.2 Hz, 3H), 0.93(s, 3H), 0.884 (s, 9H), 0.882 (s, 9H), 0.78 (s, 3H), 0.06 (s, 3H), 0.03 (s, 3H), 0.02 (s, 3H), 0.01 (s, 3H); ¹³C NMR (CDCl₃) δ 171.6, 134.2, 132.4, 128.9, 127.5, 72.7, 61.0, 57.5, 42.6, 33.2, 32.4, 31.1, 25.9, 25.8, 18.1, 18.0, 14.0, -4.0, -4.4, -5.0; IR (CHCl₃) 2955, 2929, 2857, 1735, 1472, 1256, 1096, 1072, 833, 774; HRMS (EI) m/z $(M + H^{+}) \ calcd \ 567.3360 \ for \ C_{30}H_{54}O_{4}SSi_{2}, \ found$ 567.3349. Second 1,3-trans bis silyl ether: (182 mg, 322 µmol, 93%); ¹H NMR (CDCl₃) δ 7.45–7.20 (m, 5H), 4.11 (ddd, J = 14, 7.2, 1.6 Hz, 2H), 3.74 (dd, J = 11.2, 4.8 Hz, 1H), 3.59-3.56 (m, 1H), 3.54 (d, J=8.0 Hz, 1H), 2.43–2.31 (m, 1H), 1.95–1.87 (m, 1H), 1.66–1.36 (m, 3H), 1.19 (t, J = 7.2 Hz, 3H), 0.94 (s, 3H), 0.90 (s, 9H), 0.87 (s, 9H), 0.79 (s, 3H), 0.06 (s, 3H), 0.04 (s, 3H), 0.01 (s, 3H), 0.00 (s, 3H); ¹³C NMR (CDCl₃) δ 171.7, 134.8, 131.8, 128.9, 127.3, 76.7, 72.7, 61.0, 57.8, 40.5, 35.1, 33.3, 32.7, 25.91, 25.86, 25.2, 18.5, 18.1, 18.0, 14.1, -4.1, -4.3, -5.0,-5.1; IR (CHCl₃) 2929, 2857, 1737, 1472, 1368, 1256, 1150, 909, 873, 833, 775, 691; HRMS (EI) m/z (M + H +) calcd 567.3360 for C₃₀H₅₄O₄SSi₂, found 567.3355.

1,3-trans unsaturated ester (\pm)-18. Each 1,3-trans bis silyl ether (\pm) -16 was separately subjected to the following oxidation: to a solution of bis silvl ether (182 mg, 321 µmol) in CH₂Cl₂ (7–8 mL) at 0 °C was added a solution of m-CPBA (55 mg, 321 µmol) in CH₂Cl₂ (1 mL) dropwise, maintaining the solution at less than 5°C at all times. Upon disappearance of the starting material, the reaction was quenched with saturated NaHCO₃ solution, dried over MgSO₄, and chromatographed over silica to provide two diastereomeric sulfoxides as colorless oils. From first 1,3-trans bis silvl ether above: (less polar sulfoxide, 52 mg, 90 μmol, 28%); ¹H NMR (CDCl₃) δ 7.70–7.43 (m, 5H), 3.88–3.74 (m, 3H), 3.63– 3.60 (m, 1H), 3.33 (d, J = 6.8 Hz, 1H), 2.95–2.82 (m, 1H), 1.91-1.68 (m, 3H), 1.56-1.44 (m, 1H), 0.99 (t, J = 7.2 Hz, 3H), 0.96 (s, 3H), 0.93 (s, 9H), 0.87 (s, 9H), 0.79 (s, 3H), 0.09 (s, 3H), 0.05 (s, 3H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C NMR (CDCl₃) δ 167.1, 142.8, 131.8, 128.9, 125.5, 78.3, 76.7, 72.5, 61.1, 40.6, 34.1, 33.5, 31.1, 25.92, 25.88, 25.2, 18.5, 18.1, 18.0, 13.9, -4.0, -4.3, -5.1; IR (CHCl₃) 2955, 2930, 2885, 2857, 1726, 1472, 1252, 1098, 1083, 1070, 1053, 833, 775; HRMS (EI) m/z (M+H⁺) calcd 583.3309 for C₃₀H₅₄O₅Si₂S, found 583.3313; (more polar sulfoxide, 112 mg, 192 µmol, 60%); ¹H NMR (CDCl₃) δ 7.63–7.47 (m, 5H), 4.10–3.89 (m, 2H), 3.70 (dd, J = 10.8, 4.4 Hz, 1H), 3.59 - 3.56 (m, 1H), 3.23(d, J = 8.0 Hz, 1H), 2.54–2.40 (m, 1H), 1.86–1.78 (m, 1H), 1.68-1.59 (m, 2H), 1.52-1.41 (m, 1H), 1.08 (t, J = 7.2 Hz, 3H), 0.92 (s, 3H), 0.86 (s, 9H), 0.82 (s, 9H), 0.78 (s, 3H), 0.08 (s, 6H), 0.02 (s, 3H), -0.01 (s, 3H); ¹³C NMR (CDCl₃) δ 166.2, 142.0, 131.5, 129.1, 124.7, 76.2, 75.5, 72.4, 61.2, 40.6, 34.4, 33.0, 29.7, 25.8, 25.8, 25.1, 18.5, 18.0, 14.0, -4.1, -4.4, -5.0, -5.2; IR (CHCl₃) 2955, 2929, 2857, 2855, 1732, 1472, 1255, 1094, 1057, 834, 775; HRMS (EI) m/z (M+H⁺) calcd 583.3309 for C₃₀H₅₄O₅Si₂S, found 583.3318. From second 1,3-trans bis silyl ether above: (less polar sulfoxide, 89 mg, 153 μmol, 44%); ¹H NMR (CDCl₃) δ 7.70–7.45 (m, 5H), 3.96–3.80 (m, 3H), 3.64–3.60 (m, 1H), 3.45 (d, J = 5.6 Hz, 1H), 2.95–2.86 (m, 1H), 1.86–1.72 (m, 3H), 1.01 (t, J = 7.2 Hz, 3H), 0.96 (s, 3H), 0.93 (s, 9H), 0.88 (s, 9H), 0.78 (s, 3H), 0.11 (s, 3H), 0.06 (s, 3H), 0.05 (s, 3H), 0.03 (s, 3H); ¹³C NMR (CDCl₃) δ 167.1, 142.6, 131.8, 128.9, 125.5, 77.7, 76.4, 72.6, 61.1, 40.6, 35.6, 31.0, 30.0, 25.92, 25.88, 25.2, 18.4, 18.1, 18.0, 14.0, -4.0,-4.3, -5.0, -5.1; IR (CHCl₃) 2955, 2930, 2857, 1725, 1250, 1074, 834, 775; HRMS (EI) m/z (M+H⁺) calcd 583.3309 for $C_{30}H_{54}O_5Si_2S$, found 583.3324; (more polar sulfoxide, 81 mg, 140 µmol, 40%); ¹H NMR (CDCl₃) δ 7.63–7.47 (m, 5H), 4.00–3.90 (m, 1H), 3.86– 3.72 (m, 2H), 3.58-3.55 (m, 1H), 3.18 (d, J=9.6 Hz, 1H), 2.82–2.68 (m, 1H), 2.20–2.11 (m, 1H), 1.70–1.35 (m, 3H), 1.01 (t, J = 7.6 Hz, 3H), 0.96 (s, 3H), 0.91 (s, 9H),0.90 (s, 9H), 0.81 (s, 3H), 0.08 (s, 3H), 0.04 (s, 3H), 0.03 (s, 3H), 0.02 (s, 3H); ¹³C NMR (CDCl₃) δ 165.6, 141.9, 131.1, 128.9, 124.5, 76.4, 74.5, 72.2, 61.0, 40.6, 34.9, 33.2, 29.7, 25.94, 25.91, 25.1, 18.5, 18.1, 18.0, 13.9, -3.9, -4.3,-5.17, -5.12; IR (CHCl₃) 2955, 2928, 2881, 2855, 1722, 1471, 1257, 1072, 833, 774; HRMS (EI) m/z (M+H⁺) calcd 583.3309 for $C_{30}H_{54}O_5Si_2S$, found 583.3318.

A mixture of all four aforementioned sulfoxides (334 mg, 573 µmol total) was taken up in benzene and heated

at reflux for 36 h. After removing the solvent in vacuo, silica gel chromatography (1–3% EtOAc/hexanes) provided (\pm)-18 (212 mg, 81%) as a colorless oil with the following physical characteristics: ¹H NMR (CDCl₃) δ 5.66 (br s, 1H), 4.18-4.06 (m, 2H), 3.69 (dd, J=8.0, 4.0Hz, 1H), 3.67 (dd, J=6.4, 3.6 Hz, 1H), 3.03 (dd, J = 14.4, 6.4 Hz, 1H), 2.94 (ddd, J = 14.4, 3.6, 1.6 Hz, 1H), 2.38 (ddd, J = 14.0, 4.0, 0.6 Hz, 1H), 2.22 (ddd, J = 14.0, 4.0, 1.2 Hz, 1H), 1.25 (t, J = 7.2 Hz, 3H), 0.94 (s, 3H), 0.93 (s, 3H), 0.87 (s, 9H), 0.85 (s, 9H), 0.07 (s, 3H), 0.04 (s, 3H), 0.018 (s, 3H), 0.016 (s, 3H); ¹³C NMR (CDCl₃) δ 166.4, 157.7, 116.9, 75.6, 74.9, 59.5, 41.8, 40.8, 33.4, 25.82, 25.76, 22.3, 21.1, 18.03, 17.97, 14.3, -4.1, -4.6, -5.0, -5.1; IR (CHCl₃) 2954, 2929, 2894, 2857, 1716, 1652, 1472, 1251, 1162, 1091, 1045, 835, 776; HRMS (EI) m/z (M+H+) calcd 457.3169 for $C_{24}H_{48}O_4Si_2$, found 457.3176.

1,3-trans phosphine oxide (\pm) -20. To a solution of (\pm)-18 (120 mg, 263 µmol) in Et₂O (5 mL) at-40 °C was added a solution of lithium aluminum hydride (LAH, 660 μL, 1 M in Et₂O) and the reaction mixture was warmed to 0 °C over 2 h. The reaction was quenched with a few drops of 1 N NaOH and the precipitated salts were washed with several portions of EtOAc. The combined organics were dried over MgSO₄ and the solvents removed in vacuo. Chromatographic purification of the residue over silica gel (5-10% EtOAc/hexanes) provided the corresponding allylic alcohol (104 mg, 96%) as a colorless oil: ¹H NMR (CDCl₃) δ 5.49–5.39 (m, 1H), 4.11 (d, J=6.8 Hz, 2H), 3.66-3.59 (m, 2H), 2.42-2.30 (m, 2H), 2.22-2.08 (m, 2H), 0.91 (s, 6H), 0.88 (s, 9H), 0.87 (s, 9H), 0.05 (s, 3H), 0.04 (s, 3H), 0.02 (s, 3H), 0.01 (s, 3H); ¹³C NMR (CDCl₃) δ 138.4, 124.4, 75.12, 75.09, 58.7, 41.1, 40.8, 32.9, 25.9, 25.8, 21.8, 21.7, 18.1, 18.0, -4.1, -4.2,-5.0; IR (CHCl₃) 3317 (br), 2956, 2930, 2887, 2857, 1472, 1252, 1090, 878, 836, 724; HRMS (EI) m/z $(M + NH_4^+)$ calcd 432.3329 for $C_{22}H_{46}O_3Si_2$, found 432.3338.

Following the procedure for the synthesis of (\pm) -14 above, this allylic alcohol (157 mg, 378 µmol) in THF (5 mL) was tosylated with n-BuLi (310 μL, 1.58 M in hexanes) and TsCl (97 mg, 510 µmol) in THF (1 mL), treated with potassium diphenylphosphide (KPPh₂, 1.2 mL, $0.5 \,\mathrm{M}$ in THF) at $0 \,^{\circ}\mathrm{C}$, oxidized with $\mathrm{H_2O_2}$ (200 μL, 29% aq), and purified upon passage through silica gel (30–40% EtOAc/hexanes) to provide (\pm)-20 (161 mg, 269 μmol, 71%) as a white solid: ¹H NMR (CDCl₃) δ 7.75-7.40 (m, 10H), 5.26-5.17 (m, 1H), 3.54 (dd, J = 6.8, 3.6 Hz, 1H), 3.51 (dd, J = 8.0, 4.0 Hz, 1H), 3.18– 2.97 (m, 2H), 2.31-2.22 (m, 1H), 2.06-1.96 (m, 1H), 1.81-1.72 (m, 1H), 0.84 (s, 9H), 0.83 (s, 3H), 0.81 (s, 3H), 0.809 (s, 9H), -0.026 (s, 3H), -0.029 (s, 3H), -0.04 (s, 3H), -0.05 (s, 3H); ¹³C NMR (CDCl₃) δ 139.7 (d, J=12.1 Hz), 133.0 (d, J=26.6 Hz), 132.0 (d, J=27.4)Hz), 131.7, 131.0 (t, J = 10.2 Hz), 128.4 (dd, J = 11.4, 3.8 Hz), 112.7 (d, J = 9.1 Hz), 75.2 (d, J = 1.5 Hz), 74.5 (d, J = 1.5 Hz), 40.9, 40.6 (d, J = 2.2 Hz), 33.0 (d, J = 1.5Hz), 30.2 (d, J = 69.8 Hz), 25.8, 25.7, 22.0, 21.2, 18.0, 17.9, -4.2, -5.0, -5.1; IR (CHCl₃) 3058, 2956, 2929, 2893, 2856, 1472, 1462, 1438, 1361, 1252, 1201, 1105, 1076, 1054, 877, 862, 838, 820, 804, 775; HRMS (EI) m/z (M+H+) calcd 599.3506 for $C_{34}H_{55}O_3Si_2P$, found 599.3513.

2,2-Dimethyl-19-norcalcitriol analogues (+)-8a and (+)-8b. Following the general procedure for HWE coupling found above for (-)-7a and 7b, phosphine oxide (\pm) -20 (92 mg, 0.154 mmol) in THF (1.5 mL), PhLi (95 μL, 1.68 M solution in cyclohexane/Et₂O), and (\pm) -15 (54 mg, 0.154 mmol) in THF (0.75 mL) provided 11 mg (10%) of a mixture of silyl protected analogues as a colorless oil after column chromatography (0-10% EtOAc/hexanes). This mixture was immediately taken up in EtOH (1.5 mL) and 2 mL of 20% HF in EtOH/ H₂O was added. After 48 h at rt, water was added and the mixture was extracted with EtOAc. The organic fractions were combined, washed with brine, dried over MgSO₄, and concentrated in vacuo. Silica gel chromatography (3–6% EtOH/CH₂Cl₂) provided a mixture of (+)-8a and (+)-8b as a white solid. Further purification by chiral HPLC (Daicel-Chiralpak AS, 5% EtOH/hexanes) provided the more polar analogue (+)-8a [1.0 mg, 1.5% from (+)-15] followed by analogue (+)-8b [1.9] mg, 2.8% from (+)-15] both as white solids. (+)-8a: $[\alpha] + 29$ (c 0.7, EtOH); ¹H NMR (CDCl₃) δ 6.30 (d, J = 11.0 Hz, 1H, 5.83 (d, J = 11.0 Hz, 1H), 3.68-3.60(m, 2H), 2.83-2.74 (m, 1H), 2.66 (dd, J=13.6, 3.8 Hz, 1H), 2.56 (dd, J = 13.6, 4.0 Hz, 1H) 2.35 (dd, J = 14.0, 7.6 Hz, 1H), 2.23 (dd, J = 14.0, 6.8 Hz, 1H), 1.22 (s, 6H), 1.06 (s, 3H), 1.04 (s, 3H), 0.93 (d, J = 6.4 Hz, 3H), 0.55 (s, 3H); ¹³C NMR (CDCl₃) δ 143.0, 131.5, 123.7, 115.2, 77.2, 75.4, 75.0, 71.1, 56.5, 56.3, 45.8, 44.4, 40.8, 40.6, 40.3, 36.4, 36.1, 32.7, 31.6, 29.4, 29.2, 28.9, 23.5, 22.6, 22.3, 21.2, 20.8, 20.7, 18.8, 14.1, 12.1; UV (EtOH) λ_{max} 251 nm (ε 17600). IR (CHCl₃) 3460 (br), 3021, 2962, 2928, 2854, 1602, 1458, 1377, 1261, 1097, 1031, 811; HRMS (EI) m/z (M⁺) calcd 432.3603 for $C_{28}H_{48}O_{3}$, found 432.3597. (+)-**8b**: $[\alpha]$ +21 (*c* 1.4, EtOH); ¹H NMR (CDCl₃) δ 6.29 (d, J=11.2 Hz, 1H), 5.84 (d, J = 11.2 Hz, 1H), 3.70–3.58 (m, 2H), 2.84–2.74 (m, 1H), 2.70 (dd, J = 14.0, 4.0 Hz, 1H), 2.59 (dd, J = 14.0, 3.4)Hz, 1H), 2.29 (dd, J=13.6, 9.0 Hz, 1H), 2.21 (dd, J = 14.0, 6.4 Hz, 1H), 1.22 (s, 6H), 1.07 (s, 3H), 1.03 (s, 3H), 0.94 (d, J = 6.4 Hz, 3H), 0.54 (s, 3H); ¹³C NMR (CD₃OD) δ 143.0, 131.6, 123.7, 115.1, 77.2, 75.6, 74.8, 71.1, 60.4, 56.5, 56.3, 45.8, 44.4, 40.7, 40.4, 40.3, 36.4, 36.1, 32.9, 29.4, 29.2, 28.9, 27.6, 23.5, 22.3, 20.8, 18.8, 14.2, 12.0; UV (EtOH) λ_{max} 252 nm (ϵ 33,000). IR (CHCl₃) 3021, 2927, 1602, 1457, 1377, 1032; HRMS (EI) m/z (M⁺) calcd 432.3603 for $C_{28}H_{48}O_3$, found 432.3593.

Leading to 1,3-cis compounds

1,3-cis Bis protected diol (\pm)**-17.** Each of the 1,3-cis alcohols from above [see (\pm)**-16** experimental] was protected separately in an analogous manner to the 1,3-trans alcohols to afford an identical bis silyl ether (\pm)**-17** (from first 1,3-cis alcohol: 397 mg, 701 µmol, 88%; from second 1,3-cis alcohol: 257 mg, 453 µmol, 80%) as a colorless oil: ¹H NMR (CDCl₃) δ 7.49–7.28 (m, 5H), 4.19–3.95 (m, 2H), 3.71 (d, J=12.0 Hz, 1H), 3.50–3.33 (m, 2H), 2.34–2.24 (m, 1H), 2.14–2.06 (m, 1H),

1.74–1.60 (m, 2H), 1.46–1.37 (m, 1H), 1.13 (t, J=7.2 Hz, 3H), 0.96 (s, 3H), 0.90 (s, 9H), 0.85 (s, 9H), 0.81 (s, 3H), 0.09 (s, 3H), 0.03 (s, 3H), 0.01 (s, 3H), -0.05 (s, 3H); 13 C NMR (CDCl₃) δ 172.1, 133.3, 133.0, 128.9, 128.1, 73.0, 72.8, 61.1, 54.1, 42.6, 33.2, 32.4, 31.1, 25.9, 25.8, 18.1, 18.0, 14.0, -4.0, -4.4, -5.0; IR (CHCl₃) 2955, 2929, 2857, 1738, 1472, 1256, 1150, 1074, 872, 837, 774; HRMS (EI) m/z (M+H+) calcd 567.3360 for $C_{30}H_{54}O_4SSi_2$, found 567.3366.

1,3 cis Unsaturated ester (\pm) -19. Following the oxidation procedure of (\pm) -16 above, ether (\pm) -17 (654 mg, 1.15 mmol) and *m*-CPBA (198 mg, 1.15 mmol) in CH₂Cl₂ (35 mL total) provided two diastereomeric sulfoxides as colorless oils: (less polar sulfoxide, 271 mg, 464 μmol, 40%); ¹H NMR (CDCl₃) δ 7.70–7.40 (m, 5H), 3.81-3.72 (m, 2H), 3.66 (d, J=10.4 Hz, 1H), 3.58(dd, J=10.0, 4.0 Hz, 1H), 3.43 (dd, J=10.0, 4.0 Hz,1H), 2.86–2.77 (m, 1H), 2.21–2.13 (m, 1H), 1.84 (ddd, J = 13.8, 10.0, 5.0 Hz, 1H), 1.73 (ddd, J = 14.0, 10.0, 5.2Hz, 1H), 1.52–1.44 (m, 1H), 0.98 (t, J=7.2 Hz, 3H), 0.96 (s, 3H), 0.91 (s, 9H), 0.86 (s, 12H), 0.13 (s, 3H), 0.09 (s, 3H), 0.00 (s, 3H), -0.05 (s, 3H); 13 C NMR (CDCl₃) δ 167.3, 141.8, 132.0, 128.8, 125.8, 75.9, 73.1, 72.9, 61.3, 42.0, 33.5, 32.2, 31.3, 25.9, 25.8, 25.7, 18.1, 18.0, 13.7, -4.1, -4.4, -4.98, -5.03; IR (CHCl₃) 2955, 2930, 2885, 2857, 1726, 1472, 1298, 1253, 1113, 1074, 872, 837, 775; HRMS (EI) m/z (M + H +) calcd 583.3309 for C₃₀H₅₄O₅SSi₂, found 583.3302; (more polar sulfoxide, 349 mg, 598 μmol, 52%); ¹H NMR (CDCl₃) δ 7.58– 7.47 (m, 5H), 3.84–3.74 (m, 1H), 3.71–3.60 (m, 1H), 3.48 (dd, J=11.6, 4.0 Hz, 1H), 3.40 (d, J=12.4 Hz, 1H), 3.34(dd, J=11.4, 4.2 Hz, 1H), 2.85-2.76 (m, 1H), 2.23-2.15(m, 1H), 1.86–1.75 (m, 2H), 1.41–1.32 (m, 1H), 0.99 (s, 3H), 0.89 (s, 9H), 0.85 (t, J = 7.6 Hz, 3H), 0.83 (s, 3H), 0.82 (s, 9H), 0.10 (s, 3H), 0.08 (s, 3H), -0.05 (s, 3H), -0.13 (s, 3H); ¹³C NMR (CDCl₃) δ 165.5, 140.9, 131.2, 129.0, 124.2, 72.9, 72.8, 69.7, 61.1, 43.0, 33.1, 30.6, 29.4, 25.8, 25.71, 25.66, 18.0, 17.8, 13.5, 12.4, -4.1, -4.5,-4.8, -5.1; IR (CHCl₃) 2955, 2930, 2885, 2857, 1733, 1473, 1253, 1115, 1073, 871, 837, 775; HRMS (EI) *m/z* $(M + H^{+})$ calcd 583.3309 for $C_{30}H_{54}O_{5}SSi_{2}$, found 583.3296.

A mixture of these diastereomeric sulfoxides (594 mg, 1.02 mmol total) was taken up in benzene and heated at reflux for 6 h. After removing the solvent in vacuo, silica gel chromatography (1–3% EtOAc/hexanes) provided (\pm) -19 (489 mg, 100%) as a colorless oil with the following physical characteristics: ¹H NMR (CDCl₃) δ 5.67 (t, J=1.6 Hz, 1H), 4.15 (q, J=7.2 Hz, 2H), 3.82 (ddd, J=13.6, 4.8, 1.6 Hz, 1H), 3.22 (ddd, J=11.6, 7.0,4.6 Hz, 1H), 2.39–2.30 (m, 1H), 2.13 (ddd, J = 13.2, 4.8, 1.6 Hz, 1H), 2.04–1.95 (m, 1H), 1.28 (t, J = 7.2 Hz, 3H), 0.97 (s, 3H), 0.90 (s, 9H), 0.89 (s, 9H), 0.88 (s, 3H), 0.12 (s, 3H), 0.05 (s, 3H), 0.04 (s, 3H), 0.03 (s, 3H); ¹³C NMR (CDCl₃) δ 166.2, 155.1, 116.2, 75.8, 75.5, 59.7, 42.6, 42.5, 34.3, 25.82, 25.79, 25.4, 18.02, 18.00, 14.3, 12.1, -4.1, -5.0, -5.1; IR (CHCl₃) 2956, 2930, 2894, 2858, 1717, 1652, 1472, 1362, 1253, 1241, 1161, 1140, 1112, 1079, 1051, 871, 837, 804, 775; HRMS (EI) m/z $(M+H^+)$ calcd 457.3169 for $C_{24}H_{48}O_4Si_2$, found 457.3178.

1,3-cis Phosphine oxide (\pm) -21. To a solution of (\pm) -19 $(460 \text{ mg}, 1.01 \text{ mmol}) \text{ in } \text{Et}_2\text{O} (15 \text{ mL}) \text{ at } -78 \,^{\circ}\text{C} \text{ was}$ added a solution of DIBAL-H (6 mL, 1 M in hexanes) and the reaction mixture was warmed to room temperature over 2 h. After stirring at room temperature overnight, the reaction was quenched with dilute HCl and extracted with several portions of EtOAc. The combined organics were washed with brine, dried over MgSO₄, and the solvents removed in vacuo. Chromatographic purification of the residue over silica gel provided the target allylic alcohol (194 mg, 468 μmol, 46%) as a colorless oil: ¹H NMR (CDCl₃) δ 5.46 (tt, J=6.8, 1.6 Hz, 1H), 4.14 (d, J = 6.8 Hz, 1H), 3.19 (dd, J = 11.4, 5.0 Hz, 1H), 3.13 (dd, J = 11.6, 4.8 Hz, 1H), 2.47 (ddd, J = 13.4, 4.6, 1.4 Hz, 1H), 2.28–2.18 (m, 1H), 2.08 (ddd, J = 13.2, 4.8, 1.6 Hz, 1H), 2.00–1.90 (m, 1H), 0.95 (s, 3H), 0.90 (s, 9H), 0.89 (s, 9H), 0.86 (s, 3H), 0.06 (s, 3H), 0.04 (s, 3H), 0.030 (s, 3H), 0.028 (s, 3H); ¹³C NMR $(CDCl_3)$ δ 136.9, 123.9, 76.1, 75.9, 58.6, 42.8, 41.6, 33.8, 25.8, 25.6, 18.02, 17.99, 11.9, -4.06, -4.10, -5.0; IR (CHCl₃) 3313 (br), 2956, 2930, 2887, 2857, 1472, 1463, 1362, 1255, 1078, 1050, 1006, 874, 837, 803, 774, 671; HRMS (EI) m/z (M+H⁺) calcd 415.3064 for $C_{22}H_{46}O_3Si_2$, found 415.3058.

In a manner analogous to the synthesis of (\pm) -20, this allylic alcohol (194 mg, 468 μmol), n-BuLi (385 μL, 1.58 M in hexanes), TsCl (120 mg, 632 μmol), KPPh₂ (1.4 mL, 0.5 M in THF), and H_2O_2 (250 μ L, 29% aqueous) afforded (\pm)-21 (225 mg, 80%) as an amorphous solid after silica gel chromatography (30-40% EtOAc/ hexanes): ${}^{1}H$ NMR (CDCl₃) δ 7.75–7.42 (m, 10H), 5.37-5.27 (m, 1H), 3.06 (dd, J=14.8, 7.6 Hz, 1H), 3.00(dd, J=11.0, 5.0 Hz, 1H), 2.85 (dd, J=11.4, 4.6 Hz,1H), 2.19–2.01 (m, 2H), 1.60–1.49 (m, 1H), 0.87 (s, 9H), 0.86 (s, 12H), 0.75 (s, 3H), 0.06 (s, 6H), 0.04 (s, 3H), 0.03 (s, 3H); ¹³C NMR (CDCl₃) δ 138.0 (d, J = 11.4 Hz), 132.9 (d, J = 47.9 Hz), 131.9 (d, J = 47.8 Hz), 131.8 (d, J = 5.4 Hz), 131.1 (t, J = 9.1 Hz), 128.5 (d, J = 11.4 Hz), 112.8 (d, J = 8.4 Hz), 75.7 (d, J = 3.1 Hz), 75.2 (d, J = 2.3Hz), 42.6, 41.6 (d, J=2.2 Hz), 33.7, 30.4 (d, J=69.1Hz), 25.74, 25.71, 25.4, 17.94, 17.91, 11.7, -4.0, -4.1, -4.9, -5.0; IR (CHCl₃) 3058, 2956, 2929, 2893, 2856, 1472, 1438, 1361, 1252, 1201, 1105, 1076, 1054, 877, 862, 838, 820, 804, 775; HRMS (EI) m/z (M + H $^+$) calcd 599.3506 for C₃₄H₅₅O₃Si₂P, found 599.3501.

2,2 - Dimethyl - 19 - norcalcitriol Analogues (+) - 8c and (+)-8d. Following the general procedure for HWE coupling found above for (-)-7a and 7b, (\pm)-21 (62 mg, 0.104 mmol), PhLi (62 µL, 1.68 M solution in cyclohexane/Et₂O), and (+)-15 (40 mg, 0.114 mmol) gave 32 mg (42%) of a mixture of silyl protected analogues as a colorless oil after silica gel chromatography (0-10% EtOAc/hexanes). The mixture of diastereomers was then deprotected without separation. The mixture was taken up in THF (1.5 mL) and Et₃N (35 µL) and a solution of TBAF (250 µL, 1 M in THF) added. After 48 h at room temperature, H₂O was added and the mixture was extracted with EtOAc $(3\times)$. The organic fractions were combined, washed with brine, dried over MgSO₄ and concentrated in vacuo. Silica gel chromatography (3-6% EtOH/CH₂Cl₂) provided a mixture of (+)-8c and (+)-8d as a white solid. Further purification by C-18HPLC (69% MeCN/H₂O) provided the more polar analogue (+)-8c [5.8 mg, 30% from (\pm)-21], followed by analogue (+)-8d [5.7 mg, 30% from (\pm)-21] both as white solids. (+)-8c: $[\alpha] + 94$ (c 5.8, EtOH); ¹H NMR (CDCl₃) δ 6.31 (d, J=11.2 Hz, 1H), 5.85 (d, J = 11.2 Hz, 1H), 3.53 - 3.43 (m, 2H), 2.88 - 2.75 (m, 1H),2.64-2.46 (m, 3H), 2.30 (dd, J=14.0, 6.8 Hz, 1H), 1.22(s, 6H), 1.09 (s, 3H), 1.05 (s, 3H), 0.93 (d, J = 6.4 Hz, 3H), 0.54 (s, 3H); ¹³C NMR (CD₃OD) δ 142.6, 133.0, 122.5, 117.1, 77.2, 76.7, 71.6, 58.1, 57.6, 47.0, 45.4, 42.6, 42.3, 42.0, 37.9, 37.6, 34.1, 30.9, 30.0, 28.9, 25.3, 24.7, 23.4, 22.0, 19.5, 13.1; UV (EtOH) λ_{max} 250 nm (ϵ 344,000). IR (CHCl₃) 3362, 2927, 2872, 1616, 1468, 1378, 1214, 1146, 1082, 1052, 1015, 994, 934, 880; HRMS (EI) m/z (M⁺) calcd 432.3603 for $C_{28}H_{48}O_3$, found 432.3601. (+)-8d: $[\alpha]$ +42 (c 5.0, EtOH); ¹H NMR (CDCl₃) δ 6.33 (d, J=11.6 Hz, 1H), 5.86 (d, J = 11.6 Hz, 1H, 3.58 - 3.48 (m, 2H), 2.86 - 2.77 (m, 1H),2.66-2.51 (m, 3H), 2.30 (dd, J=14.0, 5.6 Hz, 1H), 1.22(s, 6H), 1.11 (s, 3H), 1.05 (s, 3H), 0.94 (d, J = 6.8 Hz, 3H), 0.55 (s, 3H); ¹³C NMR (CD₃OD) δ 142.6, 133.0, 122.5, 117.1, 77.1, 76.7, 71.6, 58.1, 57.6, 47.0, 45.4, 42.6, 42.3, 42.0, 37.9, 37.6, 35.0, 30.9, 29.4, 29.3, 28.9, 25.3, 24.7, 23.4, 22.0, 19.5, 13.3, 12.6; UV (EtOH) λ_{max} 250 nm (ε 33,000). IR (CHCl₃) 3374, 2925, 1618, 1467, 1378, 1082, 1052, 1015, 934, 864; HRMS (EI) m/z (M⁺) calcd 432.3603 for C₂₈H₄₈O₃, found 432.3597.

PTH cell culture. Bovine parathyroid glands were obtained from a local slaughterhouse and transported to the laboratory in cold PBS. The glands were digested with collagenase as previously described⁴² and seeded at a density of 80,000 cells/cm² in DMEM–Ham's F-12 (1:1) containing 4% heat-inactivated newborn calf serum, 15 mM HEPES, 100 IU/mL penicillin, 100 µg/mL streptomycin, 5 µg/mL insulin, 2 mM glutamine, 5 µg/mL holo-transferrin and 1% non-essential amino acids. After 24 h cells were placed in medium containing 0.1% bovine serum albumin in place of the serum. Except for the initial 24 h, the cells were grown to confluency (6 days) in serum-free medium.

Analysis of parathyroid hormone secretion. Parathyroid cell cultures were prepared as described above. On the third day of culture the cells were treated with the vitamin D compounds (calcitriol or its analogues) at concentrations ranging from 10 pM to 100 nM with daily changes of the medium for 3 days. Steady state PTH secretion was determined by washing the cells three times with Dulbecco's PBS and then placing them in treatment media for 3 h. The media were collected, centrifuged at 4 °C and analyzed for PTH using CH9 antibody as described previously.⁴³

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